

Interpretive Standards for AMR Genotypes

Kat Holt | drkatholt@gmail.com

LONDON
SCHOOL of
HYGIENE
& TROPICAL
MEDICINE

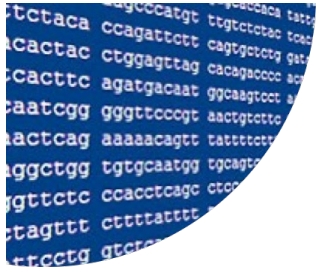


ESCMID ESGEM-AMR

amrrules.org

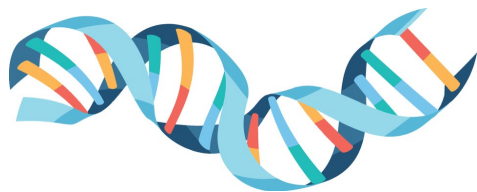


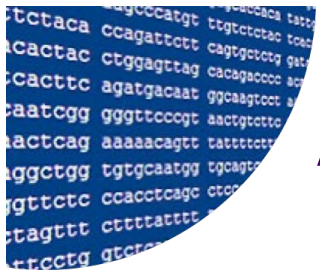
MONASH
University



Genomics for AMR

- AMR evolution & spread at different scales
- Epidemiology and transmission of AMR infections
- Outbreak investigation
- Hospital infection prevention & control (IPC)
- Sequence-based detection and diagnostics
- Public health disease surveillance
- Enviro detection & One Health AMR surveillance

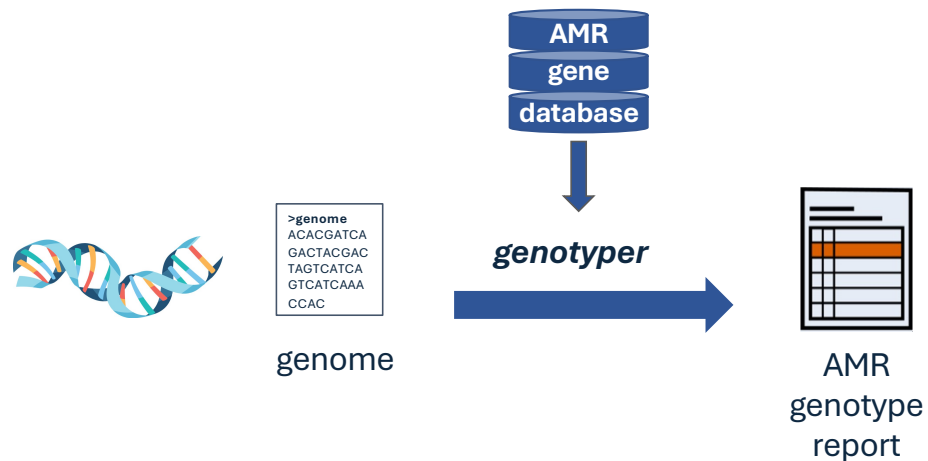




Assay: Genotyping AMR determinants in bacterial genomes

Ecosystem of bioinformatics tools and resources

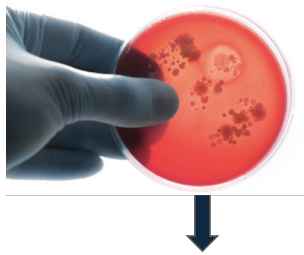
- ✓ **Databases** of AMR determinants - *NCBI, CARD, ResFinder*
- ✓ **Genotypers** to identify these in genomes – *AMRFinderPlus, CARD RGI, species-specific tools*



Missing rules for interpretation

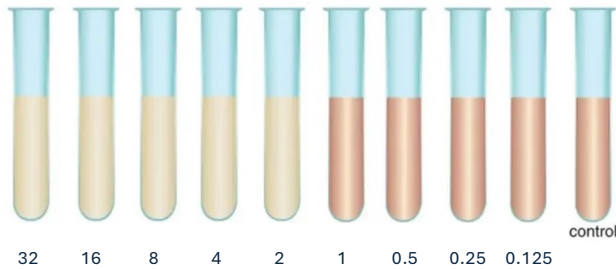
What does **variant X** in **species Y** mean for **drug Z**?

Interpretive standards for phenotypes



K. pneumoniae

Assay (microbroth dilution)



Ciprofloxacin concentration

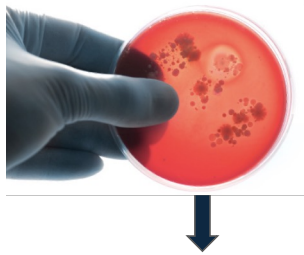
EUCAST Breakpoint Tables

Fluoroquinolones	MIC breakpoints (mg/L)		
	S ≤	R >	ATU
Ciprofloxacin, <i>Salmonella</i> spp. ¹	0.06	0.06	
Ciprofloxacin (indications other than meningitis)	0.25	0.5	0.5
Ciprofloxacin (meningitis) ²	0.125	0.125	
Pefloxacin (screen only)	NA	NA	
Delafloxacin, <i>E. coli</i>	0.125	0.125	
Levofloxacin	0.5	1	

Assay Result: MIC=2 mg/L

Interpretation: Ciprofloxacin resistant

Interpretive standards for genotypes



K. pneumoniae

Assay (WGS +AMRFinderPlus)



“expert rules”

Genotype result: *qnrS1*, *gyrA* p.Ser83Tyr

Interpretation: Ciprofloxacin resistant

Interpretation vs prediction

EUCAST interpretations (eucast.org)

S - high likelihood of therapeutic success
(using standard dosing regimen)

I - high likelihood of therapeutic success
(when exposure is increased)

R - high likelihood of therapeutic failure
even with increased exposure

Interpretation...

understand assay data, including what it means for treatment

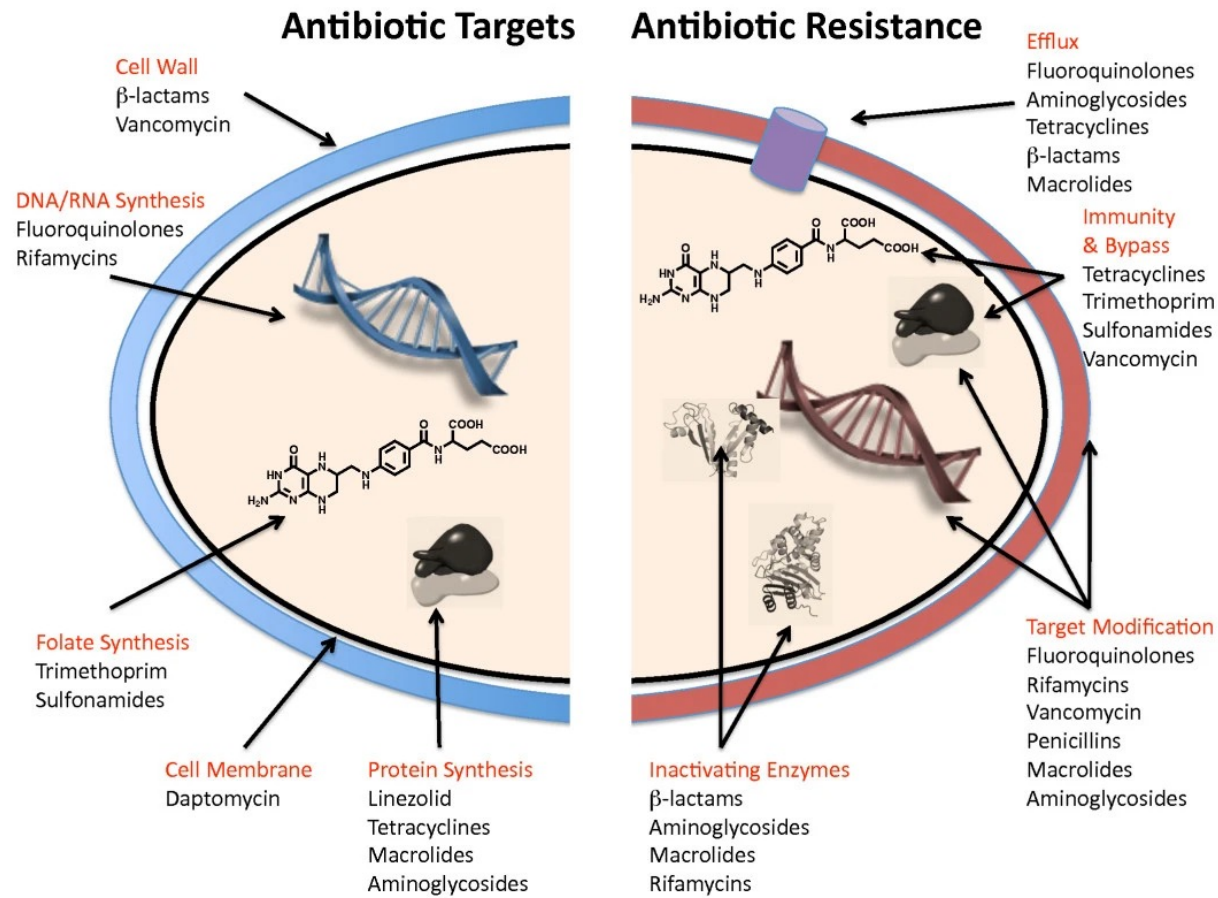
Prediction...

anticipate outcome for a given treatment, for a specific patient and context

AMR genotypes often interpreted alongside phenotypic assay measures, the goal is not to predict those assay measures

Collective knowledge of AMR genetics

Biological complexity of AMR

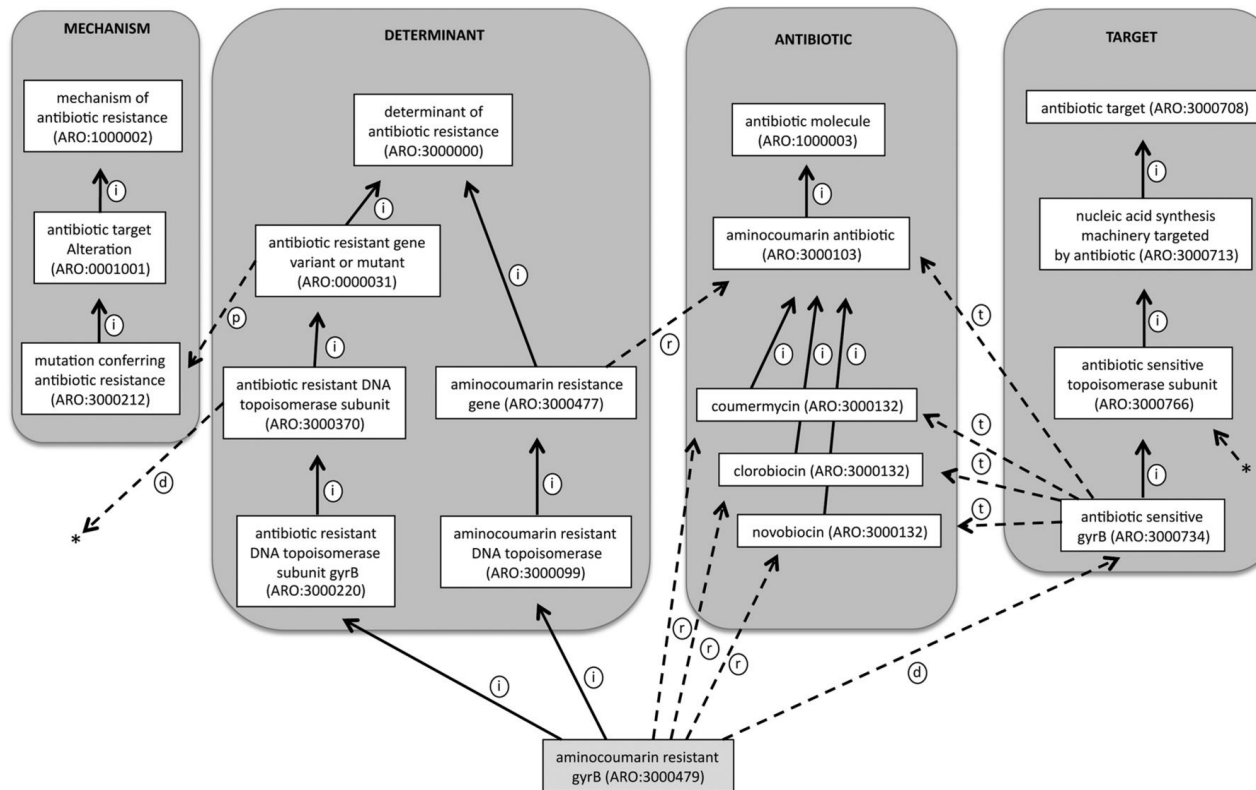


The Comprehensive Antibiotic Resistance Database

A bioinformatic database of resistance genes, their products and associated phenotypes.

7253 Ontology Terms, 5230 Reference Sequences, 1960 SNPs, 3310 Publications, 5278 AMR Detection Models

Resistome predictions: 413 pathogens, 24291 chromosomes, 2662 genomic islands, 48212 plasmids, 172216 WGS assemblies, 276270 alleles





refgene

Reference sequences:

- 8,302 AMR-associated genes
- 1,528 AMR-associated mutations

Linked PubMed IDs:

- 42% of genes
- 27% of mutations

ncbi.nlm.nih.gov/pathogens/refgene/

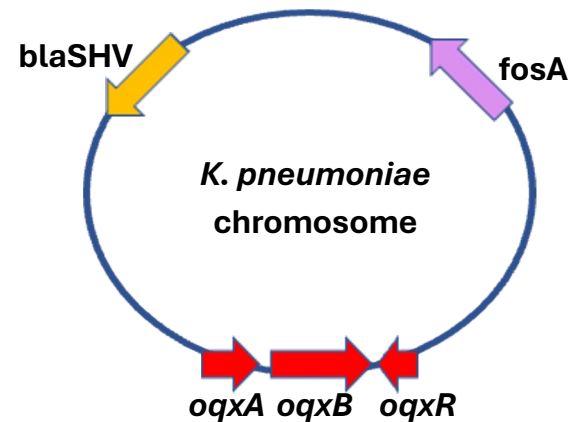
Gene family	Allele	Product name	Class	Subclass	Pubmed ref...
blaKPC	blaKPC-103	class A beta-lactamase KPC-103	BETA-LACTAM	CARBAPENEM	
blaKPC	blaKPC-104	class A beta-lactamase KPC-104	BETA-LACTAM	CARBAPENEM	35980232
blaKPC	blaKPC-105	class A beta-lactamase KPC-105	BETA-LACTAM	CARBAPENEM	
blaKPC	blaKPC-107	class A beta-lactamase KPC-107	BETA-LACTAM	CARBAPENEM	35980232
blaKPC	blaKPC-108	class A beta-lactamase KPC-108	BETA-LACTAM	CARBAPENEM	35980232
blaKPC	blaKPC-129	class A beta-lactamase KPC-129	BETA-LACTAM	CARBAPENEM	
blaKPC	blaKPC-130	class A beta-lactamase KPC-130	BETA-LACTAM	CARBAPENEM	
blaKPC	blaKPC-148	class A beta-lactamase KPC-148	BETA-LACTAM	CARBAPENEM	
blaKPC	blaKPC-193	class A beta-lactamase KPC-193	BETA-LACTAM	CARBAPENEM	
blaKPC	blaKPC-250	class A beta-lactamase KPC-250	BETA-LACTAM	CARBAPENEM	
blaKPC	blaKPC-285	class A beta-lactamase KPC-285	BETA-LACTAM	CARBAPENEM	
blaKPC	blaKPC-97	class A beta-lactamase KPC-97	BETA-LACTAM	CARBAPENEM	
blaKPC	blaKPC-10	carbapenem-hydrolyzing class A beta-lactamase KPC-10	BETA-LACTAM	CARBAPENEM	20038618
blaKPC	blaKPC-102	carbapenem-hydrolyzing class A beta-lactamase KPC-102	BETA-LACTAM	CARBAPENEM	
blaKPC	blaKPC-11	carbapenem-hydrolyzing class A beta-lactamase KPC-11	BETA-LACTAM	CARBAPENEM	22322349

AMRFinderPlus output for wildtype *Klebsiella pneumoniae*

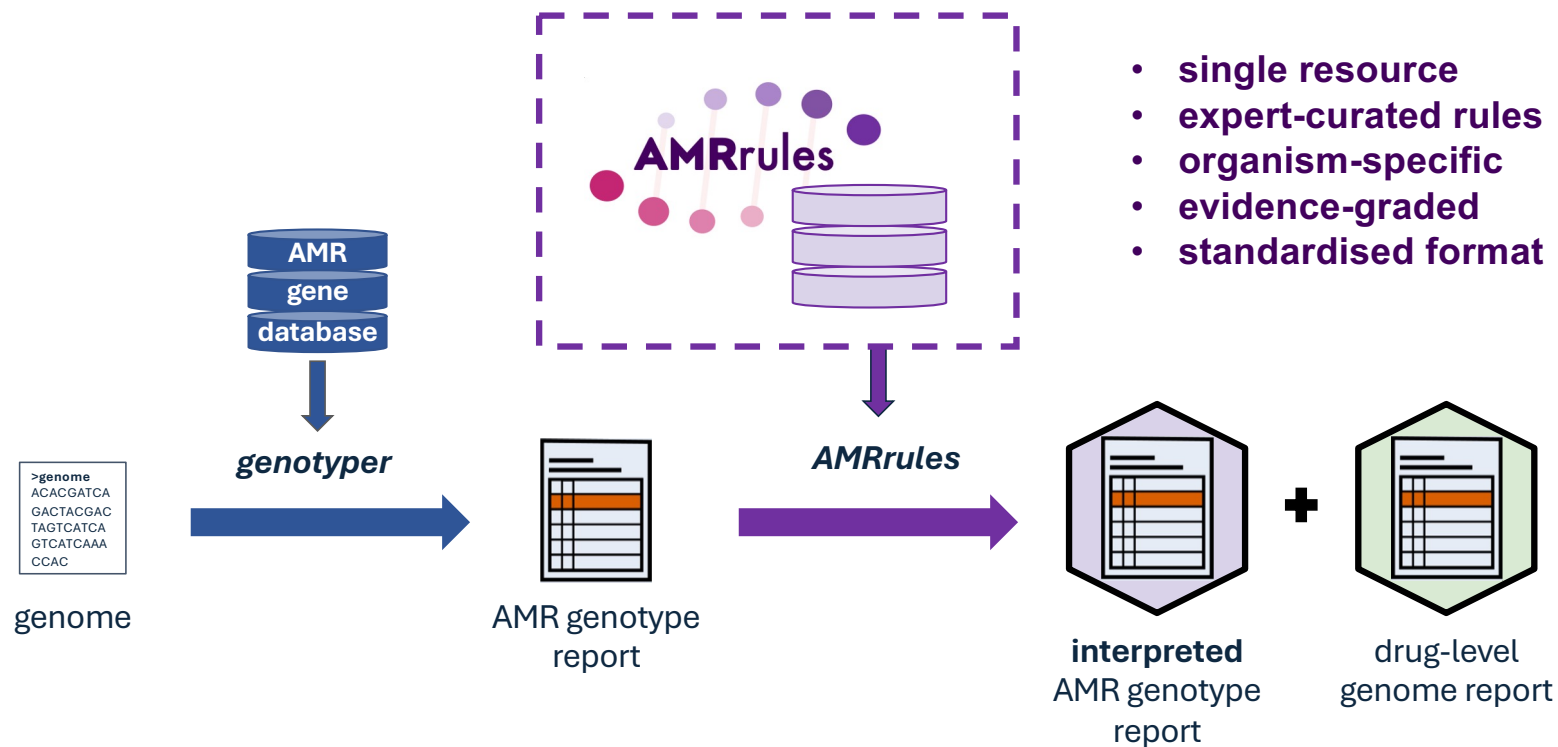
K. pneumoniae str. SGH10

Gene symbol	Class	Subclass
blaSHV-11	BETA-LACTAM	BETA-LACTAM
fosA	FOSFOMYCIN	FOSFOMYCIN
oqxA	PHENICOL/QUINOLONE	PHENICOL/QUINOLONE
oqxB19	PHENICOL/QUINOLONE	PHENICOL/QUINOLONE

all are **core genes**
in this species



Concept: expert “rules” for interpreting genotypes



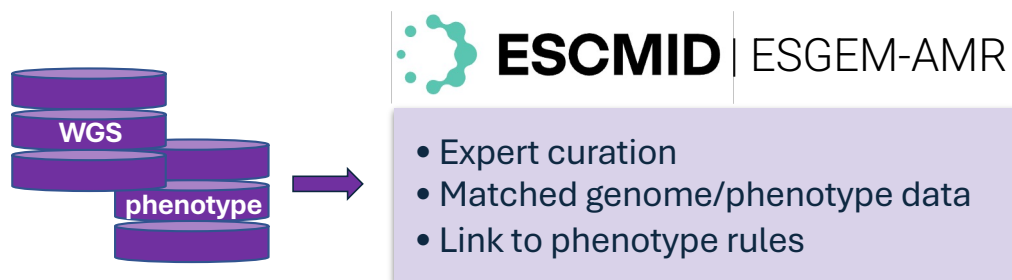


Implementation: Phase 1

- ✓ **Establish working group**
esgem-amr.amrrules.org
- ✓ **Develop rule specification and interpretation engine**
spec.amrrules.org
- ✓ **Expert subgroups define rules**
browse.amrrules.org

Working Group: ESCMID Study Group on Epidemiological Markers (ESGEM)-AMR

esgem-amr.amrrules.org



ESKAPEE / WHO AMR Priority Pathogens

Acinetobacter baumannii
Enterococcus
Enterobacter cloacae complex
E. coli/Shigella
Klebsiella pneumoniae
Staphylococcus
Pseudomonas aeruginosa
Neisseria gonorrhoeae
Salmonella
Serratia
Campylobacter spp.
Mycobacterium tuberculosis
Bordetella
Burkholderia pseudomallei
Legionella
Neisseria meningitidis
Proteus mirabilis
Shewanella
Yersinia
...more in progress

~200 members

Rule specification

spec.amrrules.org



Specify the variant to which a rule applies

spec.amrrules.org

gene	nodeID	protein accession	nucleotide accession	mutation	variation type
16S	-	-	NZ_AP019853.2: 1155759-1157299	c.1191C>T	Nucleotide variant detected
23S	-	-	NC_002946.2: 1119158-1116249	c.[2059A>G][4]	Nucleotide variant detected in multi-copy gene
blaTEM-1	blaTEM-1	WP_000027057.1	-	-	Gene presence detected
gyrA	gyrA	WP_033785346.1	-	p.Ser91Phe	Protein variant detected
mtrC	mtrC	-	NZ_CP012026.1: 1110850-1110550	c.-120C>T	Promoter variant detected
mtrD	mtrD	WP_003695678.1	-	-	Inactivating mutation detected
NG1 & NG2	-	-	-	-	Combination



RefSeq

at least ONE required to uniquely identify a reference seq

↑
HGVS

↑
hAMRonization



Specify the phenotypic interpretation

spec.amrrules.org

drug	drug class	phenotype	clinical category	breakpoint	breakpoint standard
spectinomycin	-	nonwildtype	R	MIC >64 mg/L	EUCAST v14.0 (2024)
azithromycin	-	nonwildtype	R	MIC >1 mg/L	ECOFF (January 2024)
benzylpenicillin	-	nonwildtype	R	MIC >1 mg/L	EUCAST v14.0 (2024)
ciprofloxacin	-	nonwildtype	R	MIC >0.06 mg/L	EUCAST v14.0 (2024)
azithromycin	-	nonwildtype	S	MIC <=1 mg/L	ECOFF (January 2024)
azithromycin	-	wildtype	S	MIC <=1 mg/L	ECOFF (January 2024)

CARD ARO
exactly ONE required

↑
ECOFF

Flexibility to specify interpretation using any standard

- breakpoint/s used
- source of breakpoint/s (e.g. EUCAST, CLSI, ECOFF, EUCAST Expected Resistance, PMID if no formal breakpoint)



Specify the evidence supporting a rule

spec.amrrules.org

PMID	evidence code	evidence grade	evidence limitations	curation note
2138078, 10770780	ECO:0001103 natural variation mutant evidence, ECO:0001113 point mutation phenotypic evidence	moderate	statistical geno/pheno evidence but no experimental evidence	...
20585125	ECO:0001103 natural variation mutant evidence	high	-	...
11585791, 25149062	ECO:0000005 enzymatic activity assay evidence	high	-	...
7830580, 11181352	ECO:0005027 genetic transformation evidence, ECO:0001103 natural variation mutant evidence	high	-	...
18761689	ECO:0005027 genetic transformation evidence	moderate	low clinical relevance	...
9534233	ECO:0005027 genetic transformation evidence	moderate	lacks evidence of the degree to which MIC is affected	...

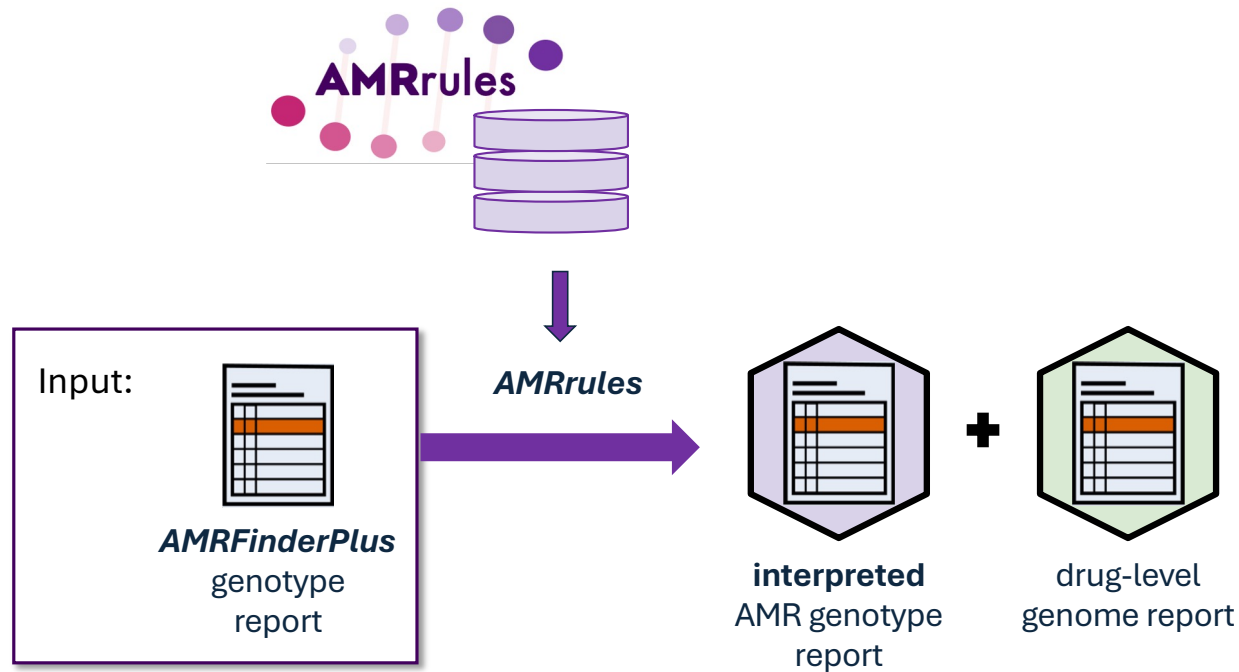
↑
PubMed

↑
Evidence & Conclusion Ontology

↑
AMRrules
(GRADE)

↑
AMRrules
(controlled terms)

Interpretation engine: AMRRules (Python package)



Jane Hawkey
Monash University
Bioinformatics Lead, AMRRules

docs.amrrules.org

amrrules.org



Output 1: Annotated genotype report



Example interpreted summary output for wildtype *Klebsiella pneumoniae* str. SGH10

Name	Gene symbol	Class	Subclass	Hierarchy node
SGH10	emrD	EFFLUX	EFFLUX	emrD
SGH10	oqxB19	PHENICOL/ QUINOLONE	PHENICOL/ QUINOLONE	oqxB19
SGH10	oqxA	PHENICOL/ QUINOLONE	PHENICOL/ QUINOLONE	oqxA
SGH10	blaSHV-11	BETA-LACTAM	BETA-LACTAM	blaSHV-11
SGH10	fosA	FOSFOMYCIN	FOSFOMYCIN	fosA5_fam



AMRFinderPlus output

(abbreviated example)



Output 1: Annotated genotype report



Example interpreted summary output for wildtype *Klebsiella pneumoniae* str. SGH10

Name	Gene symbol	Class	Subclass	Hierarchy node	ruleID	context	drug	drug class	phenotype	clinical category	evidence grade	version	organism
SGH10	emrD	EFFLUX	EFFLUX	emrD	-	-	-	-	-	-	-	1.0	-
SGH10	oqxB19	PHENICOL/ QUINOLONE	PHENICOL/ QUINOLONE	oqxB19	KPN0003	core	ciprofloxacin	-	wildtype	S	moderate	1.0	s_ <i>Klebsiella pneumoniae</i>
SGH10	oqxA	PHENICOL/ QUINOLONE	PHENICOL/ QUINOLONE	oqxA	KPN0002	core	ciprofloxacin	-	wildtype	S	moderate	1.0	s_ <i>Klebsiella pneumoniae</i>
SGH10	blaSHV-11	BETA-LACTAM	BETA-LACTAM	blaSHV-11	KPN0001	core	-	penicillin beta-lactam	wildtype	R	high	1.0	s_ <i>Klebsiella pneumoniae</i>
SGH10	fosA	FOSFOMYCIN	FOSFOMYCIN	fosA5_fam	KPN0004	core	fosfomycin	-	wildtype	S	moderate	1.0	s_ <i>Klebsiella pneumoniae</i>



AMRFinderPlus output



genotype interpretation

(abbreviated example)



Output 2: Drug-level genome summary



Example genome summary output for wildtype *Klebsiella pneumoniae* str. SGH10

drug	drug class	clinical category	phenotype	evidence grade	markers (non-S)	markers (no rule)	markers (S)	ruleIDs	combo rules	organism
ciprofloxacin	fluoroquinolone antibiotic	S	wildtype	moderate	-	-	oqxB19 (core); oqxA (core)	KPN0002; KPN0003	-	s_Klebsiella pneumoniae
(all)	penicillin beta-lactam	R	wildtype	high	blaSHV-11 (core)	-	-	KPN0001	-	s_Klebsiella pneumoniae
fosfomycin	phosphonic acid antibiotic	S	wildtype	moderate	-	-	fosA (core)	KPN0004	-	s_Klebsiella pneumoniae
(n/a)	antibiotic efflux	-	-	none	-	emrD	-	-	-	s_Klebsiella pneumoniae

interpretation

basis for it

Rule curation

browse.amrrules.org



AMRRules v1.0 1,272 rules defined in 32 species

Priorities

- ✓ Core genes in key AMR pathogens (ESKAPEE + WHO priority list)
- ✓ Phenotypes defined by EUCAST
- ✓ Compatible with AMRFinderPlus & NCBI databases

Browsing: *Klebsiella pneumoniae*

Displaying 7 row(s).

Download TSV

Columns to display

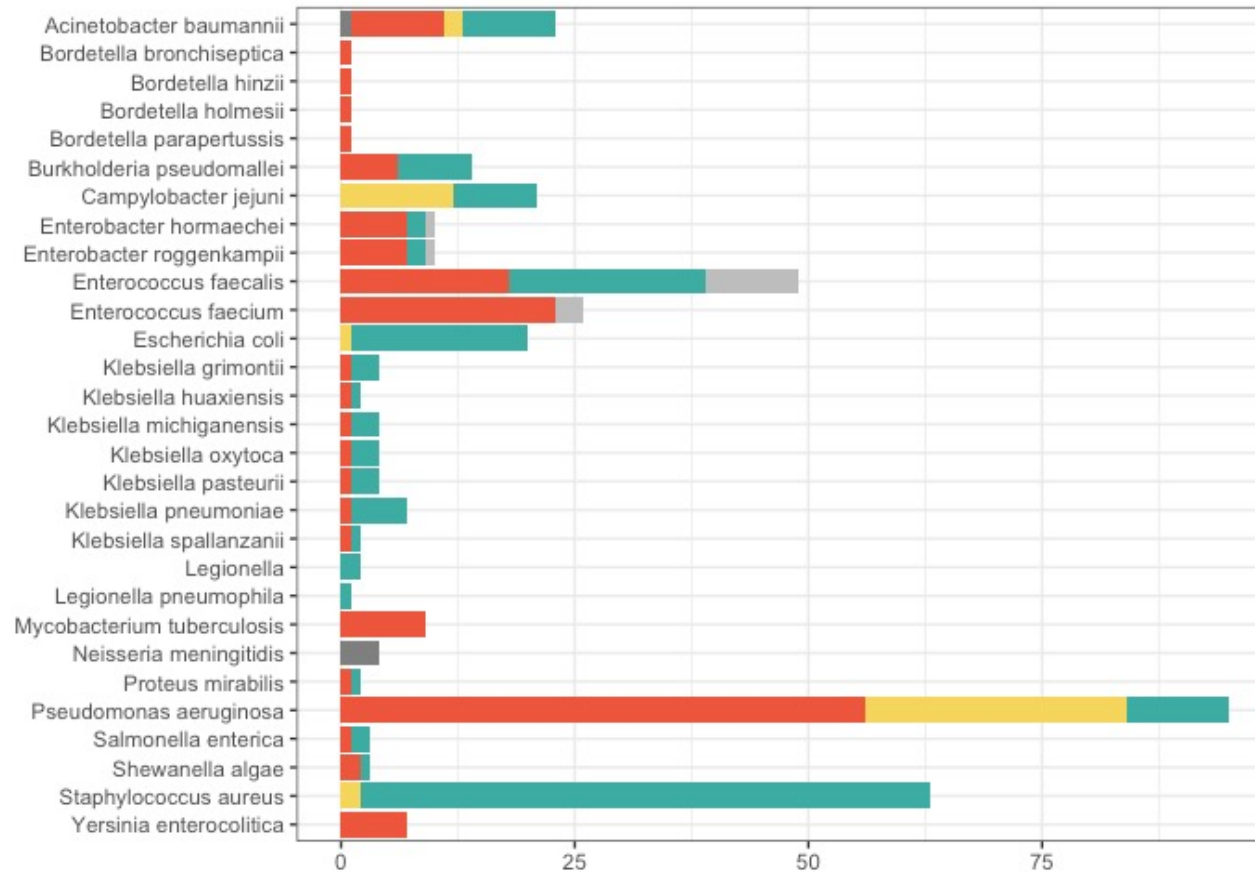
ruleID	organism	gene	mutation ⁱ	variation type ⁱ	gene context	drug	drug class	phenotype	clinical category	breakpoint standard	PMID	evidence gr...	evidence limitations ⁱ
KPN0001	Klebsiella pneumoniae	blaSHV	-	Gene presence detected	core	-	penicillin beta-lactam	wildtype	R	EUCAST Expected resistant phenotypes v1.2 (2023)	32284385	high	-
KPN0002	Klebsiella pneumoniae	oqxA	-	Gene presence detected	core	ciprofloxacin	-	wildtype	S	EUCAST v15.0 (2025)	30834112	moderate	low clinical relevance, lacks evidence for this allele
KPN0003	Klebsiella pneumoniae	oqxB	-	Gene presence detected	core	ciprofloxacin	-	wildtype	S	EUCAST v15.0 (2025)	30834112	moderate	lacks evidence for this species
KPN0004	Klebsiella pneumoniae	fosA5_fam	-	Gene presence detected	core	fosfomycin	-	wildtype	S	ECOFF (May 2025)	33128341	moderate	lacks evidence for this species
KPN0005	Klebsiella	fosA5	-	Gene	core	fosfomycin	-	wildtype	S	ECOFF (May	25441705	moderate	lacks evidence for

- ✓ All organisms
- Acinetobacter baumannii
- Bordetella bronchiseptica
- Bordetella hinzii
- Bordetella holmesii
- Bordetella parapertussis
- Bordetella pertussis
- Burkholderia pseudomallei
- Campylobacter jejuni
- Enterobacter hormaechei
- Enterobacter roggenkampii
- Enterococcus faecalis
- Enterococcus faecium
- Escherichia coli
- Klebsiella grimontii
- Klebsiella huaxiensis
- Klebsiella michiganensis
- Klebsiella oxytoca
- Klebsiella pasteurii
- Klebsiella spallanzanii
- Klebsiella pneumoniae**
- Legionella
- Legionella longbeachae
- Legionella pneumophila
- Mycobacterium tuberculosis
- Neisseria gonorrhoeae
- Neisseria meningitidis
- Proteus mirabilis
- Pseudomonas aeruginosa
- Salmonella enterica
- Shewanella
- Staphylococcus aureus
- Yersinia enterocolitica
- Yersinia pseudotuberculosis

browse.amrrules.org



Rules to interpret *presence* of core genes



393 rules

WT S (43%)

WT I (11%)

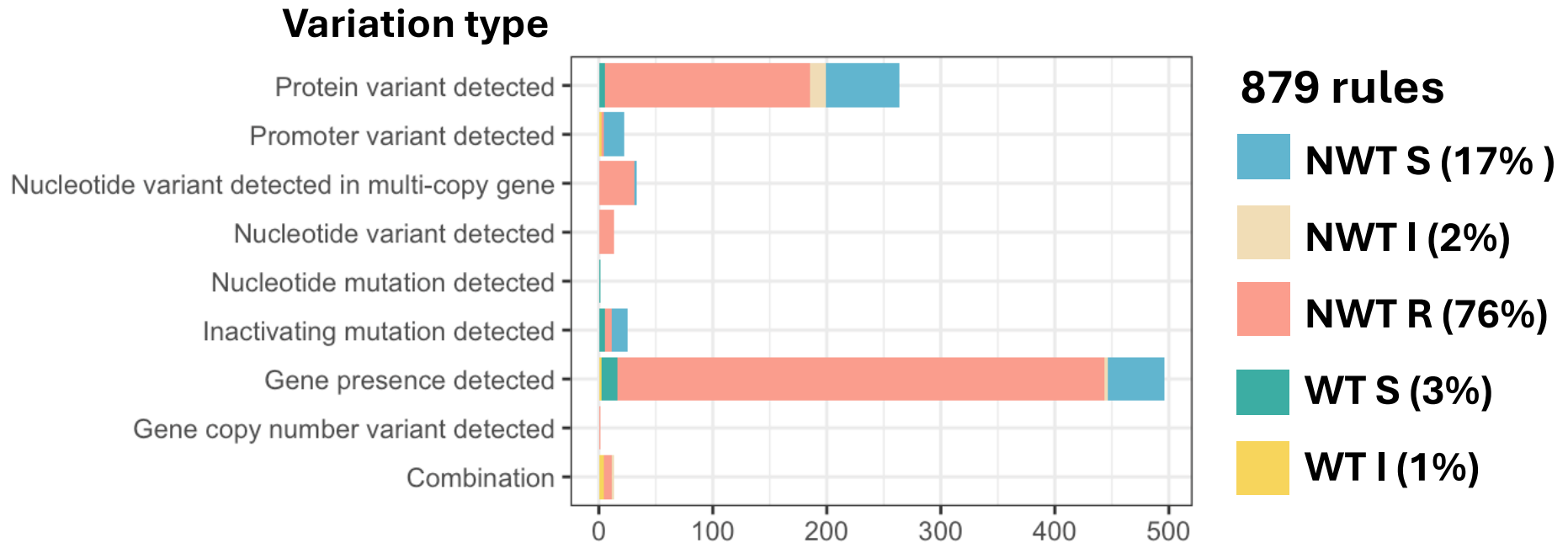
WT R (40%)

135 unique genes



Rules for acquired variants

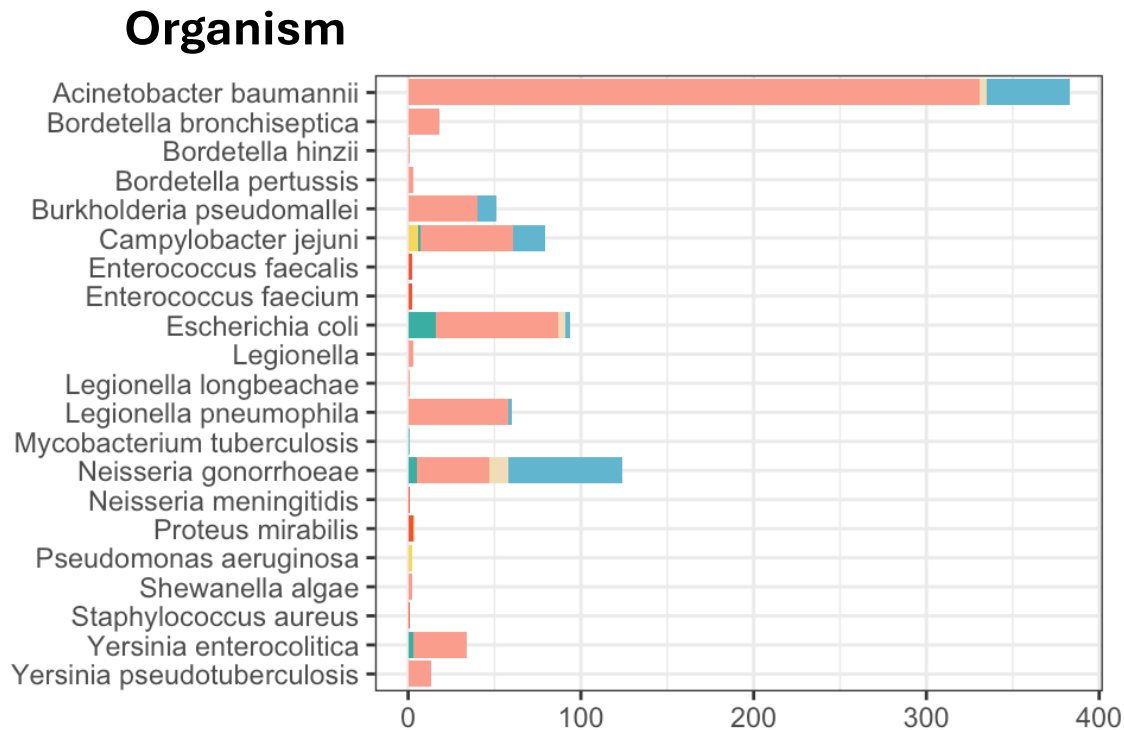
(i.e. rules other than detection of core gene presence)





Rules for acquired variants

(i.e. rules other than detection of core gene presence)



879 rules

NWT S (17%)

NWT I (2%)

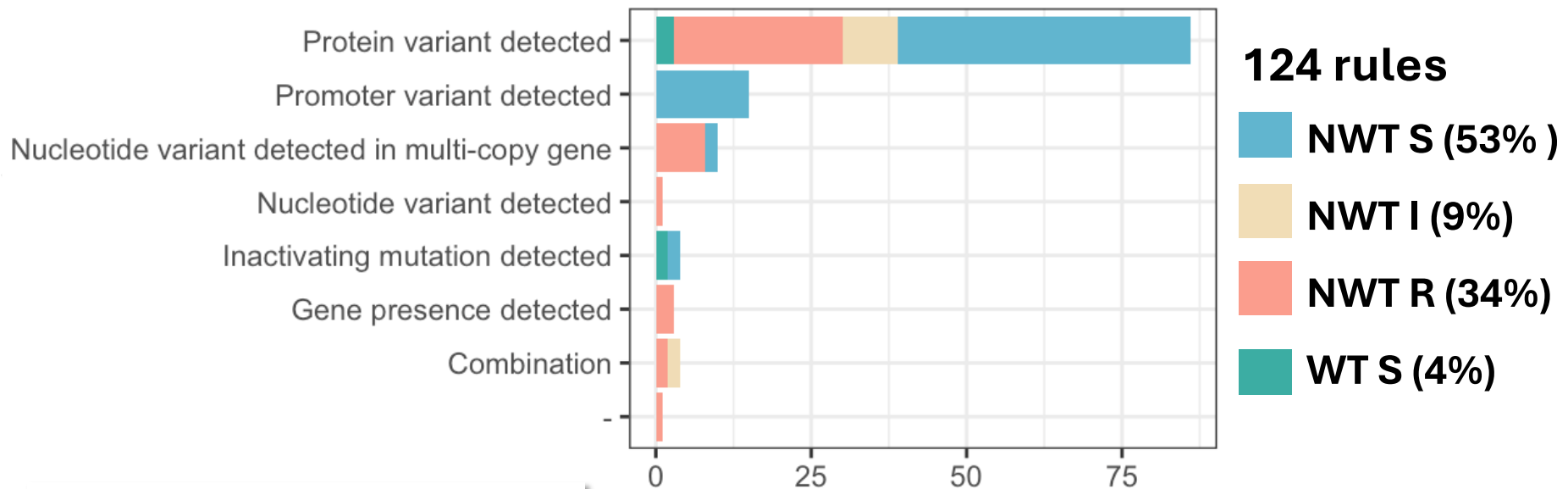
NWT R (76%)

WT S (3%)

WT I (1%)



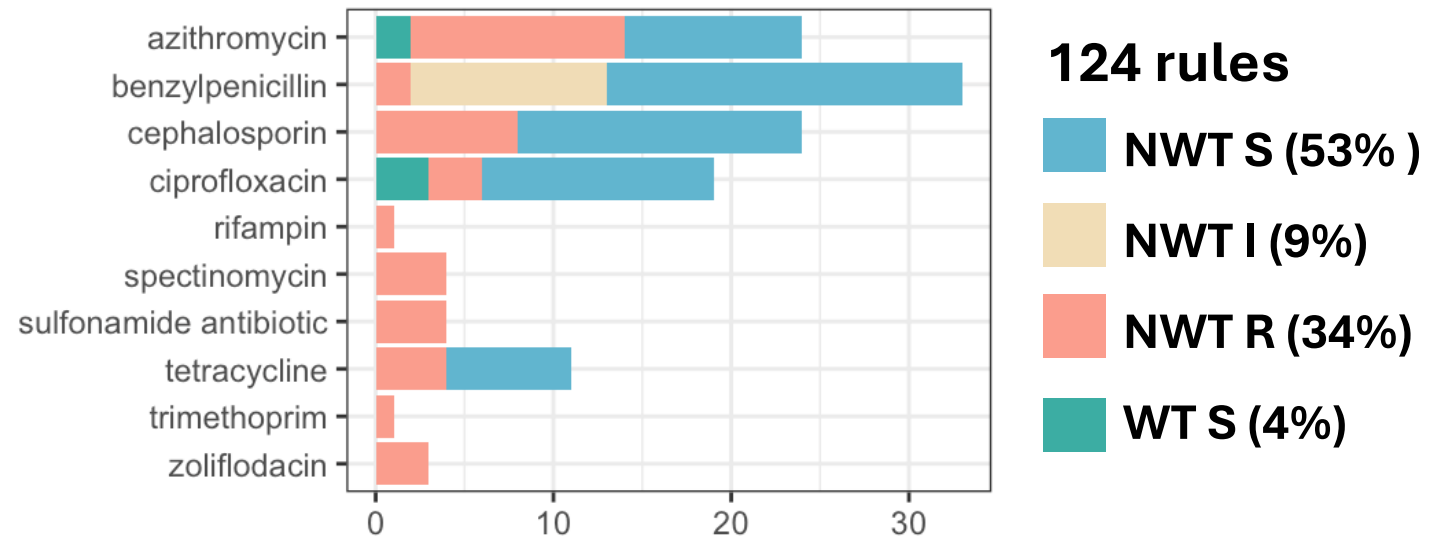
Acquired variants in *N. gonorrhoeae*



ESGEM-AMR subgroup: Leonor Sánchez Busó,
Yonatan Grad, Sheeba Manoharan-Basil, Martin
McHugh, Tatum Mortimer, Anna Roditscheff,
Faina Wehrli, Adam Witney, Raffael Frei, Daniel
Golparian, Magnus Unemo



Acquired variants in *N. gonorrhoeae*



124 rules

NWT S (53%)

NWT I (9%)

NWT R (34%)

WT S (4%)

ESGEM-AMR subgroup: Leonor Sánchez Busó,
Yonatan Grad, Sheeba Manoharan-Basil, Martin
McHugh, Tatum Mortimer, Anna Roditscheff,
Faina Wehrli, Adam Witney, Raffael Frei, Daniel
Golparian, Magnus Unemo



AMRRules interpretations for CIP in *N. gono*

Public data from EURO-GASP

Categorical concordance

95.7% Sensitivity

99.6% Specificity

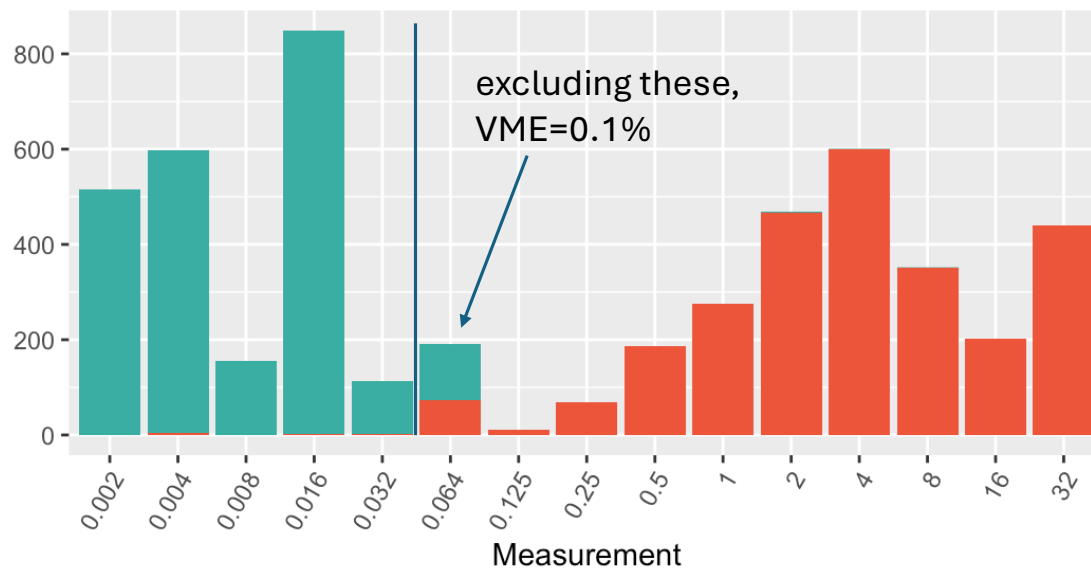
0.3% Major error (S predicted R)

4.2% Very major error (R predicted S)

[0.1% VME excluding MIC=0.64]

Observed ciprofloxacin MIC distribution

5,025 isolates



AMRRules

interpretation

WT S

NWT R



AMRRules interpretations for *A. baumannii*

Public data from EBI/CABBAGE

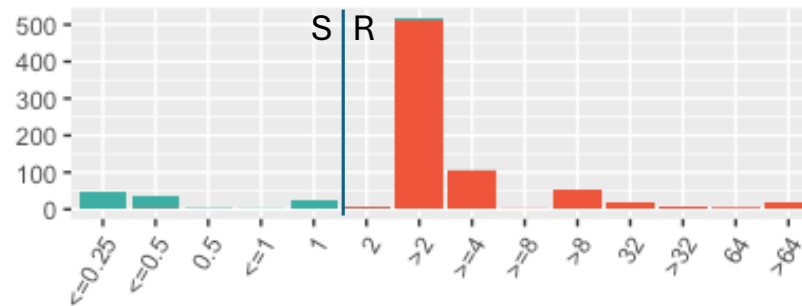
AMRRules interpretation



ESGEM-AMR subgroup:

Paul Higgins, Bogdan Iorga, Rahul Garg, Mehrad Hamidian, Priyanka Khopkar-Kate, Margaret Lam, Bruno Silvester Lopes, Ignasi Roca, Varun Shamanna, Clement Tsui, David Wareham, Valeria Bortolaia, Adrian Egli, Lucie Amoureux, Vera Manageiro, Antoine Abou Fayad, Beverly Egyir

Ciprofloxacin MIC (n=852)

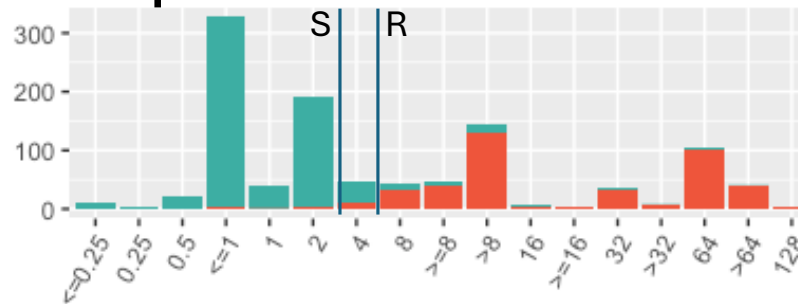


Categorical concordance

98.1% Sensitivity

99.1% Specificity

Imipenem MIC (n=1,067)



Categorical concordance

90.6% Sensitivity

97.0% Specificity



Phase 2: Quantitative rule generation

Priorities for AMRRules v2

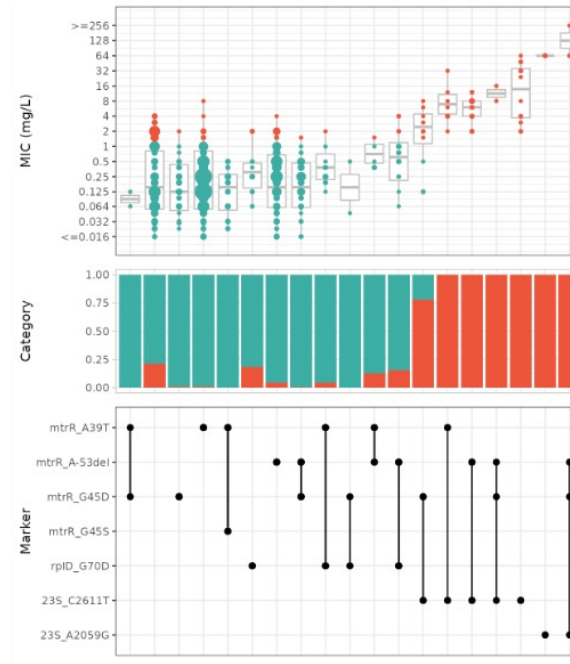
- Rules for all markers reported by AMRFinderPlus in each organism
 - ✓ Propose addition of new markers where needed
- EUCAST and CLSI phenotypic standards
- Expand AMRRules Specification to include quantitative fields
- Add support for CARD, ResFinder

Quantitative approach implemented by ESGEM-AMR



R package for AMR geno/pheno analysis

- **Import AST phenotypes**
 - NCBI, EBI, Sensititre, Phoenix, Vitek, Microscan, WHOnet
 - Re-interpret with latest breakpoints/ECOFF via AMR R pkg
- **Import AMR genotypes**
 - AMRFinderPlus, CARD RGI, ABRicate, Kleborate
- **Retrieve public data**
 - Genotypes and phenotypes from NCBI & EBI
 - Reference MIC/disk zone distributions from EUCAST
- **Format phenotype data for submission**
 - To NCBI & EBI
- **Analyse and visualise geno/pheno data**
 - Logistic regression
 - Positive predictive value
 - Assay distributions
 - Upset plots
 - Concordance metrics



Vignettes

- [Analysing Geno-Pheno Data](#)
- [Downloading geno-pheno data from NCBI and EBI](#)
- [Assessing geno-pheno concordance](#)
- [Example using large-scale regional/national surveillance data](#)
- [Example with custom stratification by isolate source](#)
- [Example with custom classification of genotype hits](#)
- [Analysing the impact of deletion variants on susceptibility](#)

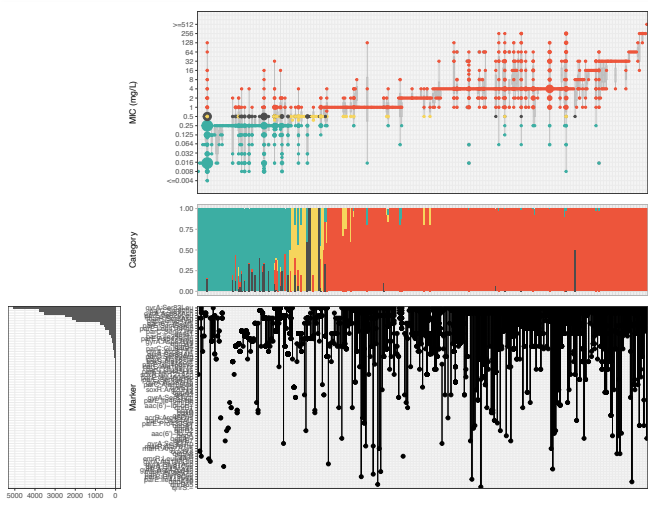
- Developers
- Matthijs S. Berends
Author, maintainer
 - Kathryn E. Holt
Author
 - Jane Hawkey
Author
 - Natacha Couto
Author
 - Ebenezer Foster-Nyarko
Author
 - Gwen M. Knight
Author
 - Zoe A. Dyson
Author
 - Kara K. Tsang
Author
 - Arjun B. Prasad
Author
 - Leonor Sánchez-Busó
Author
 - Silvia Argimón
Author
 - Dorottya Nagy
Author
 - Dominique L. Chaput
Author
 - Richard N. Goodman
Author

AMRgen.org

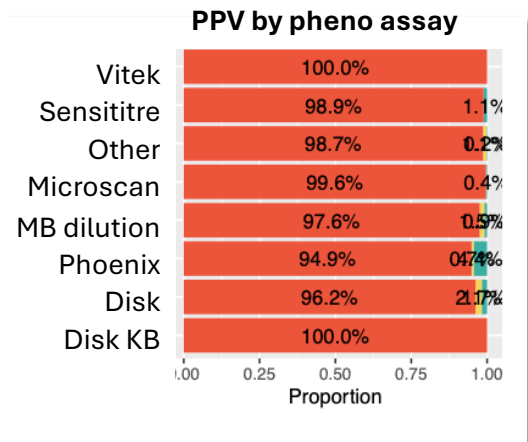


R package to define and test AMRrules

Input: geno + pheno data for a given species



ruleID	nodeID	mutation	variation type	phenotype	clinical	breakpoint
ECO1001	aac(6)-Ib-cr	-	Gene presence detected	wildtype	S	MIC <= 0.25 mg/L
ECO1002	acrR	p.Arg45Cys	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1003	emrR	p.Leu64Arg	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1004	gyrA	p.Gly81Asp	Protein variant detected	wildtype	S	disk zone >= 25 mm
ECO1005	gyrA	p.Gly81Cys	Protein variant detected	nonwildtype	S	MIC <= 0.25 mg/L
ECO1006	oqx8	-	Inactivating mutation detected	wildtype	S	MIC <= 0.25 mg/L
ECO1007	parC	p.Ala56Thr	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1008	parC	p.Ser80Ile	Protein variant detected	nonwildtype	S	MIC <= 0.25 mg/L
ECO1009	qnrA1	-	Gene presence detected	nonwildtype	S	MIC <= 0.25 mg/L
ECO1010	qnrB1	-	Gene presence detected	wildtype	S	MIC <= 0.25 mg/L
ECO1011	qnrB2	-	Gene presence detected	wildtype	S	MIC <= 0.25 mg/L
ECO1012	qnrB6	-	Gene presence detected	nonwildtype	S	MIC <= 0.25 mg/L
ECO1013	qnrB7	-	Gene presence detected	wildtype	S	MIC <= 0.25 mg/L
ECO1014	soxR	p.Gly121Asp	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1015	parE	p.Ile355Thr	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1016	parE	p.Asp475Glu	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1017	marR	p.Ser3Asn	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1018	parE	p.Ile529Leu	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1019	gyrA	p.Ser83Ala	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1020	parC	p.Ser57Thr	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1021	gyrA	p.Asp87Gly	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1022	gyrA	p.Asp87Tyr	Protein variant detected	nonwildtype	S	MIC <= 0.25 mg/L
ECO1023	soxS	p.Ala12Ser	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1024	gyrA	p.Asp87Asn	Protein variant detected	nonwildtype	S	MIC <= 0.25 mg/L
ECO1025	qnrS13	-	Gene presence detected	nonwildtype	I	MIC > 0.25 & <= 0.5 mg/L
ECO1026	qnrB19	-	Gene presence detected	nonwildtype	S	MIC <= 0.25 mg/L



Automated ciprofloxacin resistance rules for *E. coli*



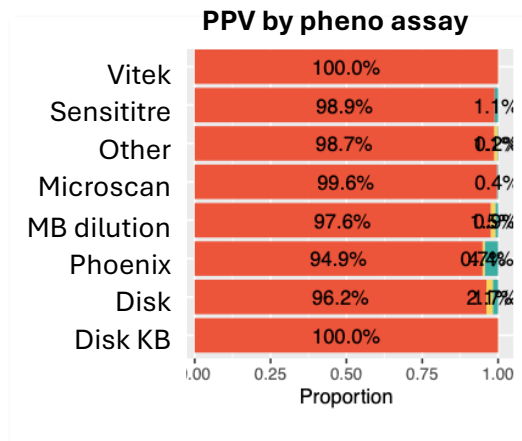
R package to define and test AMRRules

Input: geno + pheno data for a given species

➔ Propose AMRRules

ruleID	nodeID	mutation	variation type	phenotype	clinical	breakpoint
ECO1001	aac(6)-Ib-cr	-	Gene presence detected	wildtype	S	MIC <= 0.25 mg/L
ECO1002	acrR	p.Arg45Cys	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1003	emrR	p.Leu64Arg	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1004	gyrA	p.Gly81Asp	Protein variant detected	wildtype	S	disk zone >= 25 mm
ECO1005	gyrA	p.Gly81Cys	Protein variant detected	nonwildtype	S	MIC <= 0.25 mg/L
ECO1006	oqxB	-	Inactivating mutation detected	wildtype	S	MIC <= 0.25 mg/L
ECO1007	parC	p.Ala56Thr	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1008	parC	p.Ser80Ile	Protein variant detected	nonwildtype	S	MIC <= 0.25 mg/L
ECO1009	qnrA1	-	Gene presence detected	nonwildtype	S	MIC <= 0.25 mg/L
ECO1010	qnrB1	-	Gene presence detected	wildtype	S	MIC <= 0.25 mg/L
ECO1011	qnrB2	-	Gene presence detected	wildtype	S	MIC <= 0.25 mg/L
ECO1012	qnrB6	-	Gene presence detected	nonwildtype	S	MIC <= 0.25 mg/L
ECO1013	qnrB7	-	Gene presence detected	wildtype	S	MIC <= 0.25 mg/L
ECO1014	soxR	p.Gly121Asp	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1015	parE	p.Ile355Thr	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1016	parE	p.Asp475Glu	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1017	marR	p.Ser3Asn	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1018	parE	p.Ile529Leu	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1019	gyrA	p.Ser83Ala	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1020	parC	p.Ser57Thr	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1021	gyrA	p.Asp87Gly	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1022	gyrA	p.Asp87Tyr	Protein variant detected	nonwildtype	S	MIC <= 0.25 mg/L
ECO1023	soxS	p.Ala12Ser	Protein variant detected	wildtype	S	MIC <= 0.25 mg/L
ECO1024	gyrA	p.Asp87Asn	Protein variant detected	nonwildtype	S	MIC <= 0.25 mg/L
ECO1025	qnrS13	-	Gene presence detected	nonwildtype	I	MIC > 0.25 & <= 0.5
ECO1026	qnrB19	-	Gene presence detected	nonwildtype	S	MIC <= 0.25 mg/L

➔ Internal validation



ESCMID | ESGEM-AMR

Outputs curated by organism experts

- ✓ Biological and clinical sense check
- ✓ Add experimental evidence & review evidence grades
- ✓ Exploring AI-assisted biocuration

Automated ciprofloxacin resistance rules for *E. coli*



Phase 3: Routine update and review cycle

Automate updating reviews based on new data

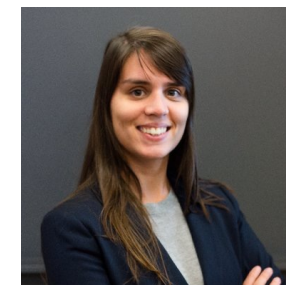
- Link to public (EBI/NCBI) and/or private (WHO/regional) geno/pheno DBs
 - EBI/NCBI, WHO/regional/national
- Expert committee for review and oversight

Interoperability

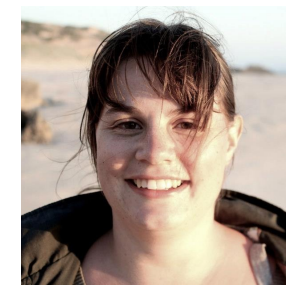
- Maintain/extend compatibility with relevant tools and standards
- Contribute to standardised approaches to AMR genomics
 - E.g. Facilitate converting AMRRules generic output into dynamic formats, applying local reporting rules, etc



ESCMID | ESGEM-AMR



Natacha Couto
Chair, ESGEM & ESGEM-AMR



Jane Hawkey
Monash University
Bioinformatics Lead, AMRRules

Organism subgroup leads contributing rules to v1.0

Leonor Sánchez Busó, Paul Higgins, Francesc Coll, Teresa Coque, Antonio Oliver, Kara Tsang, Ebenezer Foster-Nyarko, Pieter-Jan Ceysens, Conor Meehan, Birgitta Duim, Po-Yu Liu, Laurence Luu, Claire Chewapreecha, Charlotte Michel, Célia Bettencourt, Axel Hamprecht.

Members: Antoine Abou Fayad, Manal AbuOun, Dylan Adlard, Brian Alcock, Lucie Amoureux, Diego Andrey, Silvia Argimón, Sarah Baines, Kate Baker, Sangeeta Banerji, Mike Beeton, Jennifer Bender, Gherard Biffignand, Kathleen Boiten, Rémy Bonnin, Emily Bordeleau, Valeria Bortolaia, Sylvain Brisse, Ana Budimir, Rafael Cantón, Megan Carey, Luis Gustavo Carvalho Pacheco, Lina Cavaco, Leonid Chindelevitch, Chalita Chomkatekaew, Daniela M Cirillo, Iñaki Comas, Diana Costa, Chiara Crestani, Stefano De Giorgi, Ala-Eddine Deghmane, Thomas Demuyser, Deepali Desai, Ghislaine Descours, Stefanie Desmet, Sabrina Di Gregorio, Margo Diricks, Nick Duggett, Brody Duncan, Zoe Dyson, Adrian Egli, Beverly Egyir, Mohammed Elbediwi, Holly Grace Espiriu, Nicholas Feasey, Michael Feldgarden, Val Fernandez, Nancy Flountzi, Brian Forde, Philip Fowler, Raffael Frei, Ana R. Freitas, João Pedro Furlan, Teresa G. Ribeiro, Miglé Gabrielaitė, Rahul Garg, Christophe Ginevra, Stephan Goettig, Daniel Golparian, Richard Goodman, Theo Gouliouris, Yonatan Grad, Mehrad Hamidian, Dag Harmsen, Patrick Harris, Kristy Horan, Hsien-Po Huang, Po-Hsiui Huang, Alasdair Hubbard, Bogdan Iorga, Jon Iredell, Elita Jauneikaite, Sankarganesh Jeyaraj, Abdurrahman Hassan Jibril, Ulrik Stenz Justesen, Rolf Kaas, Jelalu Kemal Birmeke, Priyanka Khopkar-Kale, Béla Kocsis, Adam Komorowski, Tom Koritnik, Robert Kozak, Marcela Krutova, Margaret Lam, Fernando Lázaro Perona, Marcela Lecuit, Georgina Lewis-Woodhouse, Xena Li, Malgorzata Ligowska-Marzeta, Sam Lipworth, Iren Høyland Löhr, Bruno Silvester Lopes, Mariana López, Carla Lopez Causape, Lorena Lopez cerero, Finlay Maguire, Ramon Maluping, Vera Manageiro, Sheeba Manoharan-Basil, Elena Martinez, Luis Martinez-Martínez, Wesley Mattheu, Berends Matthijs, Andrew McArthur, Martin McHugh, Matthias Merker, Elisenda Miro, Trefor Morris, Tatum Mortimer, Alexander Moura, Karyn Mukiri, Soe Yu Naing, Jalees Nasir, Ângela Novais, Alexandra Nunes, Monica Oleastro, Precious Osadebamwen, Lisa Pählman, Benjamin Parcell, Sally Partridge, Oliver Pearse, João Perdigão, Sabine Pereyre, Romain Pogorelcnik, Arjun Prasad, Mario Ramirez, Amogelang Raphenya, Tim Read, Sandra Reuter, Ignasi Roca, Anna Roditscheff, Carla Rodrigues, Charlene Rodrigues, Assaf Rokney, John Rossen, Etienne Ruppé, Regina Russanova, Francesca Saluzzo, Ørjan Samuelsen, Derek Sarovich, Janko Sattler, Patrick Meyer Sauter, Cyril Savin, Helena Seth-Smith, Juliette Severin, Shereen Shaban Abdelkareem, Varun Shamanna, Raphael Sieber, Roberto Sierra, Carolina Silva Nodari, Leonor Silveira, Jørgen Skov Jensen, Anthony Smith, Jarraud Sophie, Nicole Stoesser, Birgit Strommenger, Tiffany Ta, Louise Teixeira-Cerdeira, Tee Keat Teoh, Clement Tsui, Gültekin Ünal, Magnus Unemo, Ana Vale, Filipa Vale, Linda Veloo, Rietie Venter, Fiona Walsh, Timothy Walsh, Yu Wan, David Wareham, Jessica Webb, Faina Wehrl, Guido Werner, Mackenzie Wilke, Adam Witney, Kelly Wyres, Basil Britto Xavier, Ting-Kuang Yeh.

esgem-amr.amrrules.org

amrgen.org

amrrules.org

Where does AMRrules fit in the ecosystem & workflows?

